



Enzymatic inactivation under high pressure processes: Models, simulation and mathematical analysis

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Modelos Matemáticos en Ciencia y Tecnología

Introduction

- The demand of **safe and minimally processed food**, prepared for immediate consumption has increased significantly in order to give service to the needs of restaurants, collective dining rooms, domestic consumption, etc.
- One of the technologies that can be used for the processing of food is **high pressure**. These treatments have the great advantage of not being based on the incorporation of additives and they allow to **avoid treatments with high temperatures** which have adverse effects on some **nutritional** and **organoleptic** properties of the food.
- Here we focus on the **modelling** and **simulation** of the effect of the combination of **high pressure** with **thermal treatments** on food, considering the **inactivation** that may take place on certain **enzymes**, as well as on the **stability** of the model.
- Due to the high computational complexity needed for solving the full models, we also consider and study a **simplified version** of them.

Mathematical model for inactivation of enzymes

The evolution of the **activity A of an enzyme** can be described by a first-order kinetic equation of the following type:

$$\frac{dA(t)}{dt} = -\kappa(P(t), T(t)) A(t), \quad (1)$$

where t is the time (min), $P(t)$ is the pressure (MPa) at time t , $T(t)$ is the temperature (K) at time t and $\kappa(P, T)$ is the **inactivation rate** (min^{-1}). Here $\kappa(P, T)$ is chosen, depending on the considered enzyme, among the following expressions:

1-

$$\kappa(P, T) = \kappa_r \exp\left(-B\left(\frac{1}{T} - \frac{1}{T_r}\right)\right) \exp(-C(P - P_r)),$$

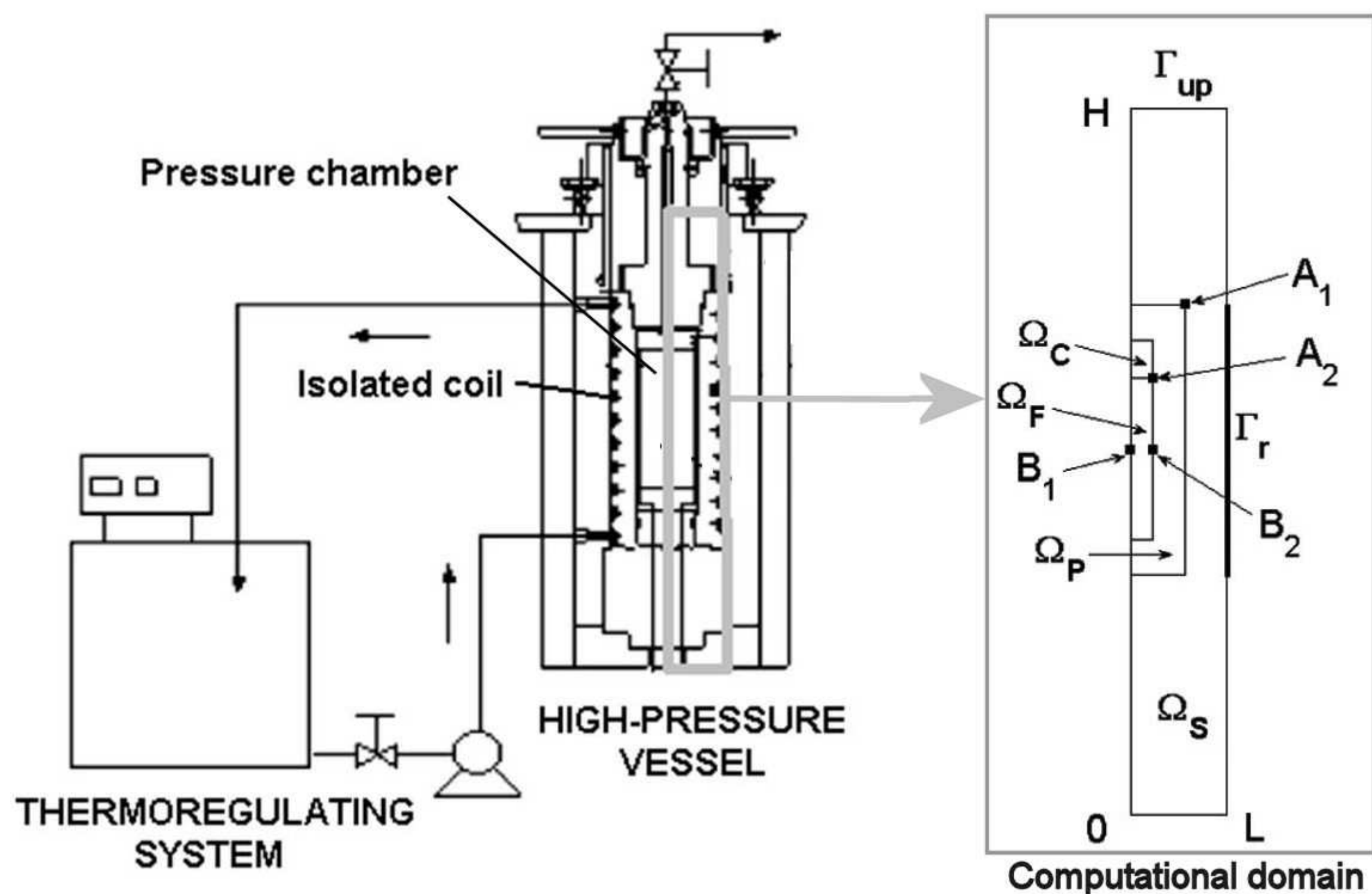
where T_r is a reference temperature (K), P_r is a reference pressure (MPa), κ_r is the inactivation rate at reference conditions (min^{-1}), B is the parameter (K) expressing the temperature dependence of κ and C is the parameter (MPa^{-1}) expressing the pressure dependence of κ .

2-

$$\kappa(P, T) = \kappa_r \exp\left[\left(\frac{-\Delta V_r}{RT}(P - P_r)\right) + \left(\frac{\Delta S_r}{RT}(T - T_r)\right) + \left(\frac{\Delta \nu}{2RT}(P - P_r)^2\right) + \left(\frac{-2\Delta \zeta}{RT}(P - P_r)(T - T_r)\right) + \left(\frac{\Delta C_p}{RT}\left(T \ln \frac{T}{T_r} - 1\right) + T_r\right)\right]$$

where $R = 8.314$ ($\text{J mol}^{-1} \text{K}^{-1}$) is the universal gas constant, ΔV_r is the volume change at reference conditions ($\text{cm}^3 \text{mol}^{-1}$), ΔS_r is the entropy change at reference conditions ($\text{J mol}^{-1} \text{K}^{-1}$), ΔC_p is the heat capacity change ($\text{J mol}^{-1} \text{K}^{-1}$), $\Delta \zeta$ is the thermal expansibility factor ($\text{cm}^3 \text{mol}^{-1} \text{K}^{-1}$) and $\Delta \nu$ is the compressibility factor ($\text{cm}^6 \text{J}^{-1} \text{mol}^{-1}$).

Considered high pressure device



ACB GEC Alstom – Instituto del Frío - CSIC.

Heat and mass transfer modelling

Considering a liquid food sample in Ω_F^* , the **governing equations** inside the whole domain Ω^* are:

$$\begin{cases} \rho C_p \frac{\partial T}{\partial t} - \nabla \cdot (k \nabla T) + \rho C_p \mathbf{u} \cdot \nabla T = \alpha \frac{dP}{dt} T & \text{in } \Omega^* \times (0, t_f), \\ \rho \frac{\partial \mathbf{u}_F}{\partial t} - \nabla \cdot \eta (\nabla \mathbf{u}_F + \nabla \mathbf{u}_F^t) + \rho (\mathbf{u}_F \cdot \nabla) \mathbf{u}_F = -\nabla p - \frac{2}{3} \nabla (\eta (\nabla \cdot \mathbf{u})) + \rho \mathbf{g} & \text{in } \Omega_F^* \times (0, t_f), \\ \rho \frac{\partial \mathbf{u}_P}{\partial t} - \nabla \cdot \eta (\nabla \mathbf{u}_P + \nabla \mathbf{u}_P^t) + \rho (\mathbf{u}_P \cdot \nabla) \mathbf{u}_P = -\nabla p - \frac{2}{3} \nabla (\eta (\nabla \cdot \mathbf{u})) + \rho \mathbf{g} & \text{in } \Omega_P^* \times (0, t_f), \\ \frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \mathbf{u}_F) = 0 & \text{in } \Omega_F^* \times (0, t_f), \\ \frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \mathbf{u}_P) = 0 & \text{in } \Omega_P^* \times (0, t_f), \\ + \text{points, boundary and initial conditions,} \end{cases} \quad (2)$$

where \mathbf{u}_F is the food velocity field, \mathbf{u}_P is the pressurizing fluid velocity field, ρ is the density (kg m^{-3}), C_p is the heat capacity ($\text{J kg}^{-1} \text{K}^{-1}$), k is the thermal conductivity ($\text{W m}^{-1} \text{K}^{-1}$), t_f (min) is the final time, \mathbf{g} is the gravity vector (m min^{-2}), η is the dynamic viscosity (Pa min), P is the pressure (Pa) applied by the equipment, p is the pressure (Pa) generated by the mass transfer inside the fluid and α is the thermal expansion coefficient (K^{-1}). * means 3D-domains obtained by **rotating** around the (0,H)-axis.

We also consider a simplified version of previous model considering the **Boussinesq approximation** (i.e. considering the incompressible Navier-Stokes equations and constant model coefficients). This approximation allows to **reduce the computational time by 3**.

Numerical tests

- Processes:** We consider two 15-minute processes:
 - **Process P_A :** For initial temperature $T_0=40^\circ\text{C}$ in Ω_S and 22°C in $\Omega \setminus \Omega_S$, a constant pressure increase in the first 305 seconds until reaching 600 MPa is considered.
 - **Process P_B :** Considering initial temperature $T_0 = 40^\circ\text{C}$ in the whole domain Ω and applying the same constant pressure increase in the first 183 seconds until reaching 360 MPa.
- Numerical Models:** The **axial symmetry** of the model allows us to work in a **2D-domain**. The heat and mass transfer model (2) is **coupled** with the kinetic equation (1). Velocity and pressure spacial discretization is based on **P2-P1 la-grange finite elements** satisfying the LBB stability condition.
- Enzymes:** We study the impact of the two processes P_A and P_B on the inactivation of three different enzymes: **Bacillus Subtilis α -Amylase (BSAA)**, **Lipoxigenase (LOX)** and **Carrot Pectin Methyl-Esterase (CPE)**.
- Sensitivity analysis:** In practice, the model coefficients are usually **approximated using experimental data**. Furthermore, due to equipment limitations, some **experimental discrepancies** could occur during the process. In order to study the impact of these errors on the enzymatic activity evolutions, we perform a **sensitivity study** of the models considering a standard deviation less than $\pm 5\%$ of the parameters.

Numerical results

Proc.	Final Temperature	Temperature evolution (C)	Enzymatic Activity Evolution (%)	Activity Reduction																												
P_A				<table border="1"> <thead> <tr> <th>Model</th> <th colspan="3">Enzyme</th> </tr> </thead> <tbody> <tr> <td></td> <td>BSAA</td> <td>LOX</td> <td>CPE</td> </tr> <tr> <td>Full model</td> <td>49%</td> <td>64%</td> <td>12%</td> </tr> <tr> <td>Boussinesq</td> <td>51%</td> <td>66%</td> <td>12%</td> </tr> <tr> <td colspan="4">Sensitivity analysis</td> </tr> <tr> <td>Minimum</td> <td>41%</td> <td>50%</td> <td>9%</td> </tr> <tr> <td>Maximum</td> <td>54%</td> <td>77%</td> <td>18%</td> </tr> </tbody> </table>	Model	Enzyme				BSAA	LOX	CPE	Full model	49%	64%	12%	Boussinesq	51%	66%	12%	Sensitivity analysis				Minimum	41%	50%	9%	Maximum	54%	77%	18%
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